Course description and objectives

BIOC63H3 provides an introduction to the scientific foundation and practice of conservation biology. It reviews ecological and genetic concepts constituting the basis for conservation, including patterns and causes of global biodiversity, the intrinsic and extrinsic value of biodiversity, the main causes of the worldwide decline of biodiversity, and approaches to save it. The course combines case studies and broad conceptual approaches to showcase the interdisciplinary nature of conservation biology, and demonstrate the social, political and economic factors that affect the discipline.

The overall goal of the course is to provide students with an introduction to both the scientific basis of modern conservation biology and the application of these principles to conservation problems around the world. After completing the course, the students should exhibit familiarity with the relevant primary and secondary scientific literature and be able to locate, summarize and synthesize information from these sources.

Instructor

Dr. Péter Molnár
Department of Biological Sciences, University of Toronto Scarborough
Office: SW567A, Phone: 416-208-2247
Email: peter.molnar@utoronto.ca
Office hours: Thu 1 – 2.30pm. If this time doesn’t work for you, send me an email to schedule a meeting. I will do my best to accommodate your schedule.

Teaching Assistants
Alexander Nascou, alexander.nascou@mail.utoronto.ca
Emily Chenery, emily.chenery@mail.utoronto.ca

Prerequisites
BIOB50 and BIOB51. These are non-negotiable.

Marks breakdown

<table>
<thead>
<tr>
<th>Component</th>
<th>Percentage</th>
</tr>
</thead>
<tbody>
<tr>
<td>Written paper on the lab project</td>
<td>35%</td>
</tr>
<tr>
<td>Quizzes about field trips (Rouge Park, Tommy Thompson Park; 2.5% each)</td>
<td>5%</td>
</tr>
<tr>
<td>Midterm exam</td>
<td>25%</td>
</tr>
<tr>
<td>Final exam</td>
<td>35%</td>
</tr>
</tbody>
</table>

Times and location

Course lecture time: Tue 4 pm – 6 pm (IC230)
Tutorial time: Thu, 9 am - 12 pm, PO101
Note: Alternate weeks for the two tutorial groups (TG1 and TG2); please consult schedule below.
## Lecture schedule & Exam dates

<table>
<thead>
<tr>
<th>Date</th>
<th>Lecture</th>
<th>Activity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sep. 6</td>
<td>1/2</td>
<td>Course introduction; Biodiversity and its value</td>
</tr>
<tr>
<td>Sep. 13</td>
<td>3/4</td>
<td>Global biodiversity patterns and processes</td>
</tr>
<tr>
<td>Sep. 20</td>
<td>5/6</td>
<td>Threats to biodiversity I: Overexploitation; habitat degradation, habitat fragmentation and habitat destruction</td>
</tr>
<tr>
<td>Sep. 27</td>
<td>7/8</td>
<td>Threats to biodiversity II: Invasive species</td>
</tr>
<tr>
<td>Oct. 4</td>
<td>9/10</td>
<td>Threats to biodiversity III: Global change</td>
</tr>
<tr>
<td>Oct. 10-15</td>
<td></td>
<td><strong>Reading week</strong></td>
</tr>
<tr>
<td>Oct. 18</td>
<td>11/12</td>
<td>Threats to biodiversity IV: Case studies tropical forests, case study oceans</td>
</tr>
<tr>
<td>Oct. 25</td>
<td>13/14</td>
<td>Conservation genetics</td>
</tr>
<tr>
<td>Oct. 27</td>
<td></td>
<td><strong>Midterm exam (lectures 1-12), both TG1 and TG2, during tutorial, 9 - 12 am</strong></td>
</tr>
<tr>
<td>Nov. 1</td>
<td>15/16</td>
<td>Approaches to conservation I: Species and populations</td>
</tr>
<tr>
<td>Nov. 8</td>
<td>17/18</td>
<td>Approaches to conservation II: Species and landscapes</td>
</tr>
<tr>
<td>Nov. 15</td>
<td>19/20</td>
<td>Approaches to conservation III: Reserve design</td>
</tr>
<tr>
<td>Nov. 22</td>
<td>21/22</td>
<td>Approaches to conservation IV: Ex situ conservation, reintroduction, managed relocation</td>
</tr>
<tr>
<td>Nov. 29</td>
<td>23/24</td>
<td>Environmental economics; Sustainability</td>
</tr>
<tr>
<td>Nov. 5 (Saturday)</td>
<td></td>
<td>--- (No tutorial this week; TTP field trip on Saturday, Nov 5 instead)</td>
</tr>
<tr>
<td>Nov. 5 (Saturday)</td>
<td></td>
<td>Field trip to Tommy Thompson Park</td>
</tr>
<tr>
<td>Nov. 10</td>
<td></td>
<td>--- (Quiz about TTP: hand in to instructor at end of field trip)</td>
</tr>
<tr>
<td>Nov. 14 (Monday)</td>
<td></td>
<td>Lab reports due on turnitin.com by 23:59h.</td>
</tr>
<tr>
<td>Nov. 17</td>
<td></td>
<td>---</td>
</tr>
<tr>
<td>Nov. 24</td>
<td></td>
<td>---</td>
</tr>
<tr>
<td>Dec TBA (exam period)</td>
<td>Final exam (lectures 13-24)</td>
<td>Final exam (lectures 13-24)</td>
</tr>
</tbody>
</table>

## Tutorials & field trips; due dates for papers & quizzes

<table>
<thead>
<tr>
<th>Date</th>
<th>Lecture or Tutorial group</th>
<th>Activity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sep. 8</td>
<td>TG1</td>
<td>Data collection: herbivore damage lab project</td>
</tr>
<tr>
<td>Sep. 15</td>
<td>TG2</td>
<td>Data collection: herbivore damage lab project</td>
</tr>
<tr>
<td>Sep. 22</td>
<td>TG1</td>
<td>Field trip to Rouge Park</td>
</tr>
<tr>
<td>Sep. 22</td>
<td>TG1</td>
<td>Quiz about Rouge Park: hand in to TA at end of field trip</td>
</tr>
<tr>
<td>Sep. 29</td>
<td>TG2</td>
<td>Field trip to Rouge Park</td>
</tr>
<tr>
<td>Sep. 29</td>
<td>TG2</td>
<td>Quiz about Rouge Park: hand in to TA at end of field trip</td>
</tr>
<tr>
<td>Oct. 6</td>
<td>---</td>
<td>---</td>
</tr>
<tr>
<td>Oct. 10-15</td>
<td></td>
<td><strong>Reading week</strong></td>
</tr>
<tr>
<td>Oct. 22</td>
<td>---</td>
<td>---</td>
</tr>
<tr>
<td>Oct. 27</td>
<td>Midterm exam (lectures 1-12), both TG1 and TG2, during tutorial, 9 - 12 am</td>
<td>--- (No tutorial this week; TTP field trip on Saturday, Nov 5 instead)</td>
</tr>
<tr>
<td>Nov. 3</td>
<td>---</td>
<td>--- (Quiz about TTP: hand in to instructor at end of field trip)</td>
</tr>
<tr>
<td>Nov. 5 (Saturday)</td>
<td>TG1 &amp; TG2</td>
<td>Field trip to Tommy Thompson Park</td>
</tr>
<tr>
<td>Nov. 5 (Saturday)</td>
<td>TG1 &amp; TG2</td>
<td>Quiz about TTP: hand in to instructor at end of field trip</td>
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</tr>
</tbody>
</table>
Field trips
There are two mandatory field trips, to (1) Rouge Park, and (2) Tommy Thompson Park. Rouge Park will happen on a Thursday during regular tutorial times with the class split into two tutorial groups; the Tommy Thompson Park trip will happen on a weekend (Sat, Nov. 5) for the entire group, cf. schedule above.

(1) Rouge Park: You are expected to travel to Rouge Park independently. If you travel by TTC, you should take bus 86A towards the zoo, and get off at the penultimate station before the zoo (Meadowvale Rd at Zoo Rd; request a stop as normally nobody gets off there; see map below). Because busses tend to run late, you need to accommodate for late bus arrivals! I strongly suggest to take a bus that is scheduled to arrive by 9:00 AM, while the field hike starts at 9:15 AM from the Rouge Valley Conservation Centre (meeting place on the map). It is your responsibility to be on time, the group will not wait for you if you are late! In order to plan your bus travel times, use TTC’s trip planner at http://www3.ttc.ca/Trip_planner/index.jsp

(2) Tommy Thompson Park: It is your responsibility to get to and from Tommy Thompson Park aka Leslie Spit Park. On Saturday, Nov. 5, we will meet at 10.00 AM at the intersection of Unwin Avenue and Leslie Street, which is the official entrance of the park. There is free parking available and I encourage you to car pool! The field trip involves a hike over a distance of approximately 10 kilometers over in part rugged terrain. We will do the trip rain, snow or shine, so please come prepared with good shoes, wind- and weather-proof clothing and a brown-bag lunch including something (warm) to drink. Due to the wind, it is ALWAYS COLDER on the spit than in the city, so make sure you are well prepared!

Attendance policy in labs and lab quizzes
In contrast to lectures, attendance will be taken in all lab sessions. Only students who attend the outdoor data collection on the herbivore damage lab project (and hence contribute to the class data set that is used in the analyses) will be able to write up the paper (and thus earn a maximum of 35% of the final course grade). Similarly, only students who participate in the field trips (Rouge and Tommy Thompson Parks) will be able to take the quiz about these field trips (and thus earn a maximum of 5% of the final course grade). If you miss any of these events due to illness or other causes beyond your control, submit, within one week of the missed lab, a written request for special consideration to the
instructor, explaining the reason for missing the event and attaching appropriate documentation, such as the official University of Toronto medical certificate (http://www.illnessverification.utoronto.ca/).

**Website**
Course information will be provided on the course website on the U of T Portal: portal.utoronto.ca. Lecture slides will also be posted here before the lecture. You will need your UTORid and your password to access the site. Please refer to instructions on how to access the course website on blackboard using the information in http://www.portalinfo.utoronto.ca. It is critical that you check the website regularly.

**Audio/video recordings**
For reasons of privacy as well as protection of copyright, unauthorized video or audio recording is prohibited. If you would like to make such recordings, for whatever reason, consult with the instructor before doing so.

**Penalty for late submission**
There will be a penalty of 10% per day for assignments received late. Weekend days count as individual days. Unless there are extenuating circumstances (e.g. medical reasons with a medical certificate), a mark of zero will be applied to assignments submitted one week late or more. Heavy workloads or malfunctioning computer equipment are not legitimate reasons for late submission. If you know ahead of time that you have a legitimate reason why you cannot hand in the assignment at the due date, let the course instructor know before the due date.

**Missed exams**
Students who miss an exam for reasons entirely beyond their control may, within one week of the missed test, submit a written request for special consideration to the instructor, explaining the reason for missing the test and attaching appropriate documentation, such as the official University of Toronto medical certificate (http://www.illnessverification.utoronto.ca/).

**Academic integrity policy**
According to Section B of the University of Toronto's *Code of Behaviour on Academic Matters*, it is an offence for students to:

- use someone else's ideas or words in their own work without acknowledging that those ideas/words are not their own with an appropriate citation and/or quotation marks, i.e. to commit plagiarism.
- include false, misleading or concocted citations in their work.
- obtain unauthorized assistance on any assignment.
- provide unauthorized assistance to another student. This includes showing another student completed work.
- submit their own work for credit in more than one course without the permission of the instructor
- falsify or alter any documentation required by the University. This includes, but is not limited to, doctor's notes.
- use or possess an unauthorized aid in any test or exam.

Violation of the Code of Behaviour on Academic Matters will force the instructor to provide a written report of the matter to the Chair/Dean/Provost, and a penalty according to the U of T’s guidelines on sanctions will be put into place.
Submission of reports to Turnitin
Students will be required to submit their lab reports to Turnitin.com for a review of textual similarity and detection of possible plagiarism. In doing so, students will allow their essays to be included as source documents in the Turnitin.com reference database, where they will be used solely for the purpose of detecting plagiarism. The terms that apply to the University’s use of the Turnitin.com service are described on the Turnitin.com web site: (http://www.portalinfo.utoronto.ca/content/turnitin)

Turnitin.com is most effective when it is used by all students; however, if and when students object to its use on principle, the course offers a reasonable offline alternative. The student will then be asked to meet with the course instructor to outline and discuss the report before its final submission to demonstrate the process of creating the report according to the academic integrity policy.

Communication policy
Students are required to regularly and often check their UTOR email to receive announcements relating to the course. To inquire about course-related issues, students are strongly encouraged to solely use their UTOR email, as hotmail or other email providers are spam-filtered on a regular basis. When emailing the instructor, please begin the subject line with “BIOC63: <subject>” to make sure emails are not overlooked. It is the responsibility of the student to make sure his or her email reaches the instructor.

The instructor will not answer any questions related to material discussed in class or during the tutorials by email (unless it is a clear yes-no answer), but the student is encouraged to ask these questions during official office hours or to schedule a meeting outside office hours by email.

Accessibility
Students with diverse learning styles and needs are welcome in this course. In particular, if you have a disability/health consideration that may require accommodations, please feel free to approach the course instructor and/or the AccessAbility Services Office as soon as possible. Enquiries are confidential. The UTSC AccessAbility Services staff (located in SW302) are available by appointment to assess specific needs, provide referrals and arrange appropriate accommodations: (416) 287-7560 or ability@utsc.utoronto.ca.

Readings
There is no required reading, but most topics introduced in the lectures are covered in the recommended course book [Groom et al. 2006. Principles of Conservation Biology. Sinauer]. The book is available at UTSC’s book store (hopefully both new and used). The course’s approach in regard to exam questions is as follows: questions will only cover material introduced in class, labs, or on the field trips. If you do not understand certain concepts, the recommended book by Groom should be consulted, but anything present in the book that is not covered in the lectures will not be on the exam.

In case a certain topic is not covered in the Groom book, the lecture material originated most likely from the primary scientific literature. In each such case, there is a reference provided on the slide along with e.g. a table or figure. This reference will help you find the article using either ISI web of science (with your UTOR ID and password, on the website of the Gerstein library; http://www.library.utoronto.ca/gerstein/) or through Google scholar (does not work in all cases).
Written Paper on Lab Project:

Herbivore damage of native and introduced plant species

1. BACKGROUND

Biological invasion is a huge contributor to the current global biodiversity crisis, second only to habitat destruction (Vitousek et al. 1997). Invasive species can have many adverse effects in their introduced range, including the outcompetition of native species in local communities. There are a number of hypotheses explaining this competitive edge of invasive species over natives, of which the “enemy release” hypothesis (Keane and Crawley 2002; Colautti et al. 2004) and the “evolution of increased competitive ability” hypothesis (Liu and Stiling 2006; Hull-Sanders et al. 2007) have received a lot of attention in recent years. At the core of both hypotheses lies the observation that invasive plant species commonly suffer less damage through herbivores and pathogens (fungi, viruses) than native plants.

Because a species’ ability to defend itself against enemies is in large part a genetically controlled trait, some groups of related species might be a priori more or less well defended than other groups. Thus, to compare the damage inflicted by enemies of just any native and non-native species might not be very meaningful and represent a comparison of “apples and oranges.” In order to compare species that are similar except for their native/exotic status, we will compare plant damage in two closely related plants, a procedure called a phylogenetically controlled approach (Agrawal and Kotanen 2003; Agrawal et al. 2005; Hill and Kotanen 2009). We will hence compare herbivore damage of a pair of native and invasive plant species within a family: the native common milkweed (Asclepias syriaca) and the invasive dog-strangling vine (Cynanchum rossicum) in the family of Asclepiadaceae.

2. DATA COLLECTION

2.1. Leaf collection

The TA or course instructor will show you the location where you will sample leaves. Each student should sample four plants of each species, thus eight plants in total. For each plant, the leaf should be collected in an unbiased and uniform way. Leaves in both species are inserted opposite each other, i.e. at each node there are two leaves (Fig. 1). For both species, identify the two leaves located in the second-highest position on a given shoot, that is, not the youngest but the second-youngest leaf pair (Fig. 1). Within this pair, collect one of them (or take the remaining leaf, if you belong to lab group 2). Make sure you tear off the entire leaf. Pack it into a plastic bag for later analysis in the lab. Be careful not to put any of the bitter, white and pressurized latex (milk) of the common milkweed into your eyes!

2.2. Analysis of leaf damage

In the lab, estimate the herbivore damage per leaf. Using the acetate paper with grid cells, essentially estimate the fraction of the damaged leaf area as the damaged leaf area divided by the whole leaf area (\(\text{fraction damaged} = \frac{\text{damaged leaf area}}{\text{total leaf area}}\)).

To do so, first check whether there is any damage at all on a given leaf (even the smallest blemish should be counted). If a leaf is spotless, score its fraction damaged leaf area as 0. If there is damage, put the leaf under the acetate paper, orienting the grid cells of the acetate paper facing down. With a sharpie, trace the outline of the leaf onto the acetate paper. If there is herbivore damage along the leaf edge, reconstruct the original outline of the leaf as closely as possible. Now count how many full and partial grid cells are contained in the traced outline of the (reconstructed) leaf on your acetate paper. In
a second step, count the full and partial grid cells of the damaged leaf area. Then divide the number of damaged grid cells by the total number of grid cells of the (reconstructed) leaf.

For the next leaf damage assessment, clean the acetate paper using rubbing alcohol and paper tissue. Repeat the counting procedure for all your sampled leaves. On a piece of paper, enter your data into a table (using the organization shown in Table 1). Upon completion, give your data to the TA. The TA will assemble a class data file and send it to everybody for analysis and write-up.

![Common milkweed](image1.png) ![Dog-strangling vine](image2.png)

**Fig. 1.** Collect one of the two leaves inserted at the second highest position per plant (arrows).

<table>
<thead>
<tr>
<th>Student name</th>
<th>Plant ID</th>
<th>Total leaf area (# grids cells)</th>
<th>Damaged leaf area (# grid cells)</th>
<th>Fraction leaf damage</th>
</tr>
</thead>
<tbody>
<tr>
<td>Jane Miller</td>
<td>MW</td>
<td>38</td>
<td>6</td>
<td>0.158</td>
</tr>
<tr>
<td>Jane Miller</td>
<td>MW</td>
<td>43</td>
<td>7.5</td>
<td>0.174</td>
</tr>
<tr>
<td>Jane Miller</td>
<td>MW</td>
<td>39</td>
<td>5.5</td>
<td>0.141</td>
</tr>
<tr>
<td>Jane Miller</td>
<td>DSV</td>
<td>26</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Jane Miller</td>
<td>DSV</td>
<td>22</td>
<td>1.5</td>
<td>0.068</td>
</tr>
<tr>
<td>Jane Miller</td>
<td>DSV</td>
<td>24</td>
<td>3</td>
<td>0.125</td>
</tr>
</tbody>
</table>
3. **DATA ANALYSIS**
Run a T-test comparing the average herbivore damage of the two species. From the analysis, retrieve the mean fraction leaf damage per species including standard errors. Use this in your figure of your report. Below, find step-by-step instructions on how to prepare the xls spreadsheet for the t-test analysis, which can be done in an online free program.

3.1. Open the spread sheet in xls

![Spreadsheet image](image1)

3.2. Sort the spread sheet in xls. From Menu, select: Data > sort

![Sort menu image](image2)
3.3. From “Sort” dialogue box: “Sort by” = “Plant”; “My list has” = Header row”. Click “OK” to continue. The data should now show up sorted in Excel.

![Sort Dialogue Box]

3.4. Data analysis. In your web browser, go to the URL: http://www.graphpad.com/quickcalcs/ttest1.cfm?Format=C
Read the website thoroughly, and consider what boxes should be selected given the data set.

![QuickCalcs Website]

3.4.1. Choose data entry format: How many rows of data do we have? More than 50, therefore choose the option, “Enter or paste up to 2000 rows.”

3.4.2. Enter data:
What labels should we choose? We are comparing between Dog strangling vine and Milkweed, so use those as labels. What Values do we enter? Since we are comparing the predation between the two plant species, we want to use data from the column “Fraction Leaf Damage” (column D). You can simply copy the rows into the “Values” section of “2. Enter data”.

3.4.3. Choose a test: What test should you choose? Click on “Help me decide” to determine the test to be used for this analysis. (Hint: “Unpaired t test”)

3.4.4. View the results: Select “Calculate now”. Your analysis is immediately calculated and returned to you on the next page. Make sure to include in your text and figure the sample sizes, standard error, t-value, degrees of freedom (df) and p-value.

4. WRITING INSTRUCTIONS
4.1. Individual subunits
The text (references, figures and tables excluded) should be 1200 – 1600 words long and consist of the following parts

Title
Abstract: max. 200 words
Key words: 6
Introduction: approx. 400 words
Methods: approx. 200 words
Results: approx. 200 words
Discussion: approx. 600 words
References (do not count towards word count of a report)

Title. Concise title, potentially containing the main finding of your study.

Abstract. The abstract should explain to the general reader why the research was done and why the results should be viewed as important. It should be able to stand alone; the reader should not have to get any information from the main paper in order to understand the abstract. The abstract should provide a brief summary of the research, including the background, purpose, methods, results, and
major conclusions. Do not include literature citations in the abstract. Avoid long lists of common methods or lengthy explanations of what you set out to accomplish. The primary purpose of an abstract is to allow readers to determine quickly and easily the content and results of a paper. The following breakdown works well: general introduction of the relevance of the biological question (1-2 sentences), objectives of the study (1-2 sentences), outline of the methods (1-2 sentences), results (1-2 sentences), conclusion (1 sentence).

Key words. List 6 key words. Each key word should be useful as an entry point for a literature search if your report were to be published.

Introduction. A brief Introduction describing the paper's significance, which should be intelligible to a general reader that does not have a background in this particular area. The Introduction should state the reason for doing the research, the nature of the questions or hypotheses under consideration, and essential background. The introduction is the place where you can show the reader how knowledgeable you are with a given field, without being too lengthy. Close the introduction with your main hypothesis/question(s).

Methods. The Methods section should provide sufficient information to allow someone to repeat your work. A clear description of your experimental design, sampling procedures, and statistical procedures is especially important.

Results. Results generally should be stated concisely and without interpretation. Present your data using figures and tables; guide your reader through them.

Discussion. The discussion section should explain the significance of the results. Distinguish factual results from speculation and interpretation. Avoid excessive review. Structure your discussion as follows. 1. First paragraph - restate your major findings concisely and relate them to the literature. 2. Compare your findings with those in other studies; examine why differences may have occurred. 3. Discuss the problems that might have been present to influence your findings. 4. End with an outlook paragraph that could discuss, e.g. how future studies could build on your research, whether any new interesting questions arose, or the relevance of your findings for invasive species management.

References. Use the correct format (see “Formatting the references” section below). You should search for and read related studies beyond those cited in the overview of the lab; your report should list at least 12 references, of which at least 9 should be new (i.e. not included in the lab instructions).

4.2. Formatting your report, writing tips
Use the formatting style of the journal of “Ecology.” It might seem tedious to you to have to follow the many rules the journal prescribes, but adhering to one style makes a paper more organized, increases readability, and bad formatting typically is a sign that the contents are also of sub-par quality.

Formatting of species names. When mentioning a species in English, also provide the Latin name at first mention. Latin names have to be in italics and the first time a Latin name is mentioned, the genus name (first part of the official binomial name) has to be spelled out; later on it can be abbreviated, such as in the following example: “Common milkweed, *Asclepias syriaca*, is a hermaphroditic perennial common to Southern Ontario. The leaves of *A. syriaca* are toxic to cattle.”
**Formatting of references.** In the body of the text, references to papers by one or two authors in the text should be in full, e.g. Agrawal and Kotanen (2003) show blablabla. Or: Blablabla (Agrawal and Kotanen 2003). If the number of authors exceeds two, they should always be abbreviated; e.g. Agrawal et al. (2005) show blablabla. Or: Blablabla (Agrawal et al. 2005). If providing more than one reference in brackets, the order should be chronological with the oldest first and the younger ones later. In the case of two studies from the same year, the order should be alphabetical. E.g. Blablabla (Zuk 1963; Korpelainen 1998; Stehlik and Barrett 2005, 2006; Stehlik et al. 2008).”

All references cited (and read by you!) in the main text should be included in “Literature cited.” References should be in alphabetical order and their formatting should follow the format exemplified below.

**Citing articles in scientific journals:**

**Citing whole books:**

**Citing individual articles/chapters in books (if the individual chapters have different authors than the book):**

**Citing a webpage (should be the exception, so avoid as much as possible; cite a paper or book instead):**

**Formatting of tables.** Tables (if present) should NOT be inserted in your text, but follow, one table per page, after your Literature cited. Give a brief description what the table is about (table caption) and introduce the parameters stated in the table in a text inserted above the table. The description should be self-explanatory, thus the reader should not be forced to read the main body of text in order to understand the message of a table. Each column and row in the table should be labeled (with units if necessary). If mentioning a species name, provide the spelled out Latin name (in italics). In the table, round numbers to two meaningful digits.

**Formatting of figures.** The design of a figure should clearly convey a major result, thus scale your data appropriately. Label all axes with sufficiently large font and meaningful labels. Keep it simple; do not use unnecessary elements such as 3D diagrams if not absolutely necessary as based on the data structure. Similarly as tables, figures should NOT be inserted in your text, but follow, one figure per page, after your tables. Give a brief description what the figure is about (figure caption) and introduce the parameters stated in the figure in a text inserted above the figure. The description of the figure should be self-explanatory; the reader should not be forced to read the main body of text in order to understand the message of a figure. Also, each axis in a plot should be labeled (with units) and each bar
in a bar chart should be labeled. If mentioning a species name, provide the spelled out Latin name (in italics).

**References to tables and figures in the text.** In your text, refer to figures as follows: ‘In the spring, temperatures are higher than in the winter (Fig. 1).’ Or: ‘Figure 1 shows that temperatures are higher in the spring than in the winter’. In your text, refer to tables as follows: ‘In the spring, temperatures are higher than in the winter (Table 1).’ Or: ‘Table 1 shows that temperatures are higher in the spring than in the winter’.

**Formatting of statistical references.** In the text, the results of a statistical test should be cited in parentheses, in support of a specific statement. When mentioning the result of a statistical test, always provide the mean values, standard errors or confidence intervals, the test statistic, degrees of freedom, and the p-value. Format your text according to the following example. “There was a significant difference in the frequency of flowering between low and high elevation sites, with greater bias among low than high elevation populations (average flowering frequency: low elevation = 0.93, SE = 0.01; high elevation = 0.78, SE = 0.02; χ² = 35.04, df = 1, p < 0.0001).”

**Miscellaneous.** Avoid quotations - paraphrase your sources (while making sure you are not plagiarizing) and provide the appropriate reference.

5. **GENERAL GRADING SCHEME FOR REPORTS**

When writing the report, you should consider the criteria and grading scheme that will used to evaluate your report.

5.1. **Information content (30%)**

This portion of the grade reflects whether or not you have presented and adequately discussed all of the relevant information. This includes background information on the topic being addressed, as well as the information you have gathered (or should have gathered). Specifically, do not forget to include all relevant statistical result parameters, statistical and other tables, data figures and the written explanation of the results. Also make sure you have cited the adequate number of required articles.

27-30: All of the relevant information was included and discussed adequately.
24-26: One of the pieces of information was not included or discussed adequately.
20-23: One of the most important pieces of information was not included or discussed adequately.
10-20: Two or more of the most important pieces of information were not included or discussed adequately.
<10: Little of the important information was included or discussed.

5.2. **Interpretation and persuasiveness (30%)**

This portion of your grade reflects whether or not you interpreted the information correctly and provided persuasive arguments to support your interpretation. Specifically, does your reasoning make sense on its own and also in the light of the published literature, with which you compare your results?

27-30: All of the relevant information was interpreted correctly, and the arguments were very persuasive.
24-26: Most of the information was interpreted correctly, and the arguments were persuasive.
20-23: One of the important pieces of information was not interpreted correctly, or some of the arguments were not persuasive.
10-20: Two or more important pieces of information were not interpreted correctly, and some of the arguments were not persuasive
<10: Little of the information was interpreted correctly, and few of the arguments were persuasive.

5.3. Clarity of writing & Structure of the document (20%)
This portion of the grade reflects whether you followed the general structure of scientific articles as outlined in 4.1, and whether or not you wrote your sentences and paragraphs clearly. In particular, regarding structure, is the paper clearly and appropriately separated into Introduction, Methods, Results, Discussion etc, or are there structural issues, such as results presented in Methods (rather than in Results), interpretation of results in the Results section (rather than in the Discussion), etc? Regarding writing clarity, do you avoid overly long sentences? Are your paragraphs succinct and mostly dealing with one major line of reasoning each? Do your paragraphs preferably start with an introductory sentence and end with a strong summarizing statement? Do you use scientific terms correctly?

17-20: Very clear writing and flawless structure
13-16: Some unclear sentences and/or some structural issues
9-12: Several unclear sentences and/or several structural issues
5-8: Many unclear sentences and/or many structural issues
<5: Few clear sentences, pervasive structural issues

5.4. Formatting (10%)
This portion of the grade reflects whether or not you formatted your report well. This includes the overall structure, the references, and the figures and tables (see instructions above).

9-10: The entire report was formatted correctly, and looked very professional.
7-8: The report was formatted correctly, and looked fairly tidy.
5-6: There were a few formatting errors.
3-4: There were several formatting errors.
<3: There were many formatting errors.

5.5. Spelling, grammar and punctuation (10%)
This portion of the grade reflects whether or not you used correct spelling, grammar and punctuation.

9-10: There were no errors in spelling, grammar, or punctuation
8-9: There were a few minor errors
7-8: There were several minor errors, or a few major errors
5-7: There were several major and minor errors
<5: There were many errors