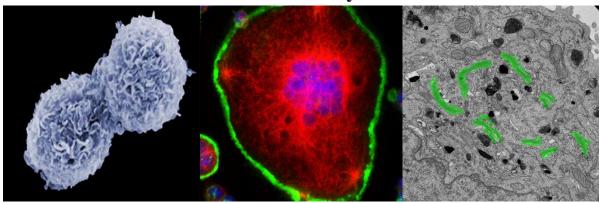
BIOD23: Special Topics in Cell Biology

Professor Rene Harrison 2017 Fall Term Syllabus



A lecture/seminar/discussion class on contemporary topics in Cell Biology. Students will explore the primary literature becoming familiar with experimental design and methodologies used to decipher cell biology phenomenon. Student seminars will follow a series of lectures and journal club discussions.

Lectures:

MW-264, Mon 1-3 p.m.

MW-110, Wed 12-1 p.m.* (*not every week- see schedule on page 4).

Textbook:

None. Dr. Harrison's PowerPoint presentations and journal articles will be supplied on blackboard as PDF files for students to download and bring to class.

Exams:

2 Exams:

- Midterm worth 25% (short and long answer questions) 2 hours
- Final exam worth 20% (short and long answer questions) 2+ hours

Office Hours:

Office: SW 541-A

Office Hours: Tuesdays 11-1 p.m., or by appointment.

e-mail: harrison@utsc.utoronto.ca

Blackboard Resources:

- Outline of PowerPoint presentations will be uploaded onto blackboard prior to class.
- Students should print and bring outlines to class to take notes on additional details.

TA:

Reuben Philip: reuben.philip@mail.utoronto.ca

Course Prerequisites:

Prerequisites:

BIOC12H [or IMCB08H plus IMCC03H (for Industrial Microbiology students only)]

Grading Scheme:

Assignments (2)	20%
Seminar	20%
Midterm	25%
Final	20%
Participation	5 %
Extracurricular	10%

100%

Assignments (20%):

Two (2) Assignments will be given. Assignments must be handed in at the <u>beginning</u> of class the day the articles are being discussed. Late assignments will be graded as <u>zero</u>. Journal articles and assignments will be put on blackboard 1-2 weeks before the discussions.

Midterm (25%):

The midterm will cover the topics/journal articles and assignments covered up until the date of the exam. The format will be short and long answer questions.

Seminar (20%):

Students (in pairs) will give a 25-minute seminar to the class. Students can choose from a recent paper (in the last 5 years) from PubMed that utilizes a microscopy technique to answer a cell biology problem. The entire paper does not need to be centered around the microscopy technique, but instead uses it for some of the data to help obtain important results. Students must decide on a paper and email the paper to Dr. Harrison for approval by October 20th at the latest. Presentation dates will be assigned on a first-come, first-serve basis. Students will present a 10-minute background on the specific microscope technology and then a 15-minute discussion on the major experiments and findings in the article using this technique. A 5-10-minute question period will follow.

Presenting students must provide a <u>3-page summary</u> (double-spaced) of the technique and research article to the other students at the end of term (<u>December 4th</u>) for the final exam.

Examples of microscopy techniques include: LSM Confocal, Spinning Disk Confocal, TIRF, FRAP, FRET, Multi-photon, Smartphone Microscopy, AFM, SIM, STORM, Optogenetics, etc.

Final (20%):

The final will include the journal articles discussed in the student seminars, particularly the 3-page summaries written by the students. For this reason, it is very important the 3-page summaries are clear and comprehensive.

Participation (5%):

Participation will be graded according to class attendance and questions asked during student seminars.

Extracurricular Activities (Chose ONE, 10% of final grade):

Students will choose ONE of the following assignments which will be worth 10% of the final grade. Specific opt-in dates are outlined below for each assignment.

A) Creative Cell Project (10%):

Students (individually or in pairs) can focus their artistic flair(s) on cell biology. Projects can include but are not limited to: computer animations for BIOB10, music videos, paper mache, cell poetry/ cell haiku, organelle stuffed animals, baked goods, your own crazy idea... Students that are interested in this must email Dr. Harrison their potential creative ideas (and partner if applicable) by October 9th. Creative projects will be presented at show-and-tell on November 6th in class.

B) Live Cell Imaging (10%):

Students will spend a Friday evening and a Saturday (tentatively October 20th and 21st) performing live cell phagocytosis imaging of transfected macrophages. Students MUST complete the Laboratory Biosafety Training certification to do this. Biosafety training is usually 2 weekday mornings in late September- dates TBD. Please see this link for more information: http://www.ehs.utoronto.ca/services/biosafety/training.htm. Full attendance at the biosafety course is mandatory for this assignment, so consider your schedules before signing up for this. Students must opt-in by sending Dr. Harrison an email by September 18th, if they want to sign up for Biosafety, and confirm training completion by October 9th. The cell imaging will be done in SW-422, and limited to 16 students max.

Schedule

DATE	TOPIC	RELEVANT PAPERS
Sept. 6 (Wed)	Class Introduction	
11	Cell Biology Electron Microscopy Techniques/ Phagocytosis	Articles 1a, 1b and 1c
13 (Wed)	No class	
18	Electron microscopes & tissue culture demo	* meet in SW 541-A
25	Journal Article Discussion#1 **Assignment#1 due	Articles 1a, 1b and 1c
27 (Wed)	Optional: Applying for grad school	
Oct. 2	Cell Biology Fluorescent Techniques/ Phagocytosis	Articles 1c, 2a and 2b
9-13	READING WEEK	
16	Fluorescent microscope / flow cytometry and cell staining/transfection demo	* meet in SW 541-A
23	Journal Article Discussion#2 **Assignment#2 due	Articles 1c, 2a and 2b
Oct.30	MIDTERM (in class)	
Nov. 6	Show and Tell day - Bone cells and Microgravity (Rene) - Creative Projects Due	
8 (Wed)	Student Seminars	
13	Student Seminars	
15 (Wed)	Student Seminars	
20	Student Seminars	
22 (Wed)	Student Seminars	
27	Student Seminars	
29 (Wed)	Student Seminars	
Dec. 4	Student Seminars *3-pagers due	
Date TBA	FINAL EXAM	

^{**}Disclaimer: The above schedule, policies, procedures, and assignments in this course are subject to change in the event of extenuating circumstances.